# ADHP

Cat. No.:	HY-101880
CAS No.:	119171-73-2
Molecular Formula:	C <sub>14</sub> H <sub>11</sub> NO <sub>4</sub>
Molecular Weight:	257.24
Target:	Fluorescent Dye
Pathway:	Others
Storage:	4°C, protect from light
	* In solvent : -80°C, 6 months; -20°C, 1 month (protect from light)

# SOLVENT & SOLUBILITY

	Preparing Stock Solutions	Solvent Mass Concentration	1 mg	5 mg	10 mg
		1 mM	3.8874 mL	19.4371 mL	38.8742 ml
		5 mM	0.7775 mL	3.8874 mL	7.7748 mL
		10 mM	0.3887 mL	1.9437 mL	3.8874 mL

DescriptionADHP is a fluorogenic peroxidase substrate ( $\lambda_{ex}$ =530 nm, $\lambda_{em}$ =590 nm).In VitroTo obtain the parameters K <sub>m</sub> and k <sub>cat</sub> for Compound I, two independent methods are used. Initially, the oxidation of ADHP using the injector functionality built-in to the fluorescence plate reader is studied. The auto-injector dispenses the H <sub>2</sub> O <sub>2</sub> to initiate the reaction, as a means of generating a set of progress curves. Analysis for MPO-mediated oxidation of ADHP gives a K <sub>m</sub> of 31±4 µM and the k <sub>cat</sub> of 186± 6 s <sup>1</sup> . The k <sub>obs</sub> also increases over the experimental range of ADHP concentrations from 1 to 80 µM and for the converse experiment holding substrate constant over 3 to 45 nM MPO. The apparent second order rate constant obtain from the slope of k <sub>obs</sub> against ADHP concentration K <sup>app</sup> on is 2.1±0.2 mM/s <sup>[1]</sup> .	BIOLOGICAL ACTIVITY				
using the injector functionality built-in to the fluorescence plate reader is studied. The auto-injector dispenses the H <sub>2</sub> O <sub>2</sub> to initiate the reaction, as a means of generating a set of progress curves. Analysis for MPO-mediated oxidation of ADHP gives a K <sub>m</sub> of 31±4 µM and the k <sub>cat</sub> of 186± 6 s <sup>1</sup> . The k <sub>obs</sub> also increases over the experimental range of ADHP concentrations from 1 to 80 µM and for the converse experiment holding substrate constant over 3 to 45 nM MPO. The apparent second order rate	Description	ADHP is a fluorogenic peroxidase substrate ( $\lambda_{ex}$ =530 nm, $\lambda_{em}$ =590 nm).			
MCE has not independently confirmed the accuracy of these methods. They are for reference only.	In Vitro	using the injector functionality built-in to the fluorescence plate reader is studied. The auto-injector dispenses the $H_2O_2$ to initiate the reaction, as a means of generating a set of progress curves. Analysis for MPO-mediated oxidation of ADHP gives a $K_m$ of $31\pm4 \mu$ M and the $k_{cat}$ of $186\pm6$ s <sup>1</sup> . The $k_{obs}$ also increases over the experimental range of ADHP concentrations from 1 to 80 $\mu$ M and for the converse experiment holding substrate constant over 3 to 45 nM MPO. The apparent second order rate constant obtain from the slope of $k_{obs}$ against ADHP concentration $K^{app}_{on}$ is $2.1\pm0.2 \text{ mM/s}^{[1]}$ .			

# PROTOCOL

### Kinase Assay <sup>[1]</sup>

ADHP, 4-ABAH, 2-ABAH, 4-BAH, 4-FBAH, 4-NBAH, 4-TFMBAH, 3-DMABAH, NaN<sub>3</sub> and isoniazid are dissolved in DMSO and subsequently diluted into assay buffer. The final concentration of DMSO in the reaction is less than 0.5 % (v/v), which does not affect fluorescence of the oxidized ADHP product 7-hydroxyl-3H-phenoxazin-3-one (resorufin). Reactions of ADHP (20 μ

റ

HC

ΟH



M) are incubated with MPO (2.8 nm) in assay buffer and initiated by the addition of 1/10th volume  $H_2O_2$  from a serial dilution basin. To determine the effect that the simplest benzoic acid hydrazide inhibitor or its analog 4-TFMBAH has on the heme catalytic ability of MPO, MPO (1.2  $\mu$ M) is incubated for 10 min with different concentrations of BAH inhibitor (0, 0.025, 0.25, 2.5, 12.5 and 25 mM) with ADHP (40  $\mu$ M) and timing of the reaction is measured following addition of  $H_2O_2$  (20  $\mu$ M) ADHP. All reactions are measured in assay buffer at room temperature. Samples of 20  $\mu$ L are added to non-reducing sample loading buffers, and then loaded without prior heating and resolved by 4-15% gradient SDS-polyacrylamide gel electrophoresis<sup>[1]</sup>. MCE has not independently confirmed the accuracy of these methods. They are for reference only.

### REFERENCES

[1]. Jiansheng Huang, et al. Ordered Cleavage of Myeloperoxidase Ester Bonds Releases Active site Heme Leading to Inactivation of Myeloperoxidase by Benzoic Acid Hydrazide Analogs. Arch Biochem Biophys. 2014 Apr 15; 548: 74–85.

#### Caution: Product has not been fully validated for medical applications. For research use only.

Tel: 609-228-6898Fax: 609-228-5909E-mail: tech@MedChemExpress.comAddress: 1 Deer Park Dr, Suite Q, Monmouth Junction, NJ 08852, USA