



# **Screening Libraries**

**Proteins** 



# **Product** Data Sheet

# Collagenase IV, Clostridium histolytica

Cat. No.: HY-E70005D CAS No.: 9001-12-1 MMP Target:

Pathway: Metabolic Enzyme/Protease

Powder -20°C Storage: 3 years

> In solvent -80°C 6 months

> > -20°C 1 month

Collagenase IV, Clostridium histolytica

# **SOLVENT & SOLUBILITY**

In Vitro

H<sub>2</sub>O: 50 mg/mL (Need ultrasonic)

# **BIOLOGICAL ACTIVITY**

## Description

Collagenase, Type IV (EC 3.4.24.3) is a microbially derived matrix metalloproteinases (MMPs) and zinc peptidase. Collagenase, Type IV breaks down collagens 1, 3, 7, 8, 10, gelatin, proteoglycans, aggrecan<sup>[1]</sup>.

# In Vitro

Type IV collagenase is recommended for islet primary cell isolation.

### Preparation of storage solution

- 1. Add 1 mL Hank's Balanced Salt Solution (HBSS) with calcium and magnesium directly to 1 g vial of Collagenase. Vortex gently to ensure complete dissolution, and prepare a stock solution of 100 mg/mL (100X stock solution).
- 2. Filter sterilize 100X stock solution using a 0.22 μm filter with a low protein binding filtration unit. Use immediately or dispense into aliquots and store at -20°C to -5°C protected from light.
- 3. Thaw on ice prior to use. Commonly used concentrations for tissue and cell dispersion are 0.5-2.5 mg/mL and for cartilage digestion are 1-2 mg/mL, but the optimal working concentration required needs to be determined based on specific experimental conditions or by referring to the appropriate literature.

### Dissociate Tissue

- 1. Mince tissue into 3-4 mm pieces with a sterile scalpel or scissors.
- 2. Wash the tissue pieces several times with HBSS containing calcium and magnesium.
- 3. Add sufficient HBSS with calcium and magnesium to submerge tissue. Add collagenase to required working concentration.
- 4. Incubate at 37°C for 4-18 hours. Increased efficiency is obtained using a rocker platform and supplementing the digest with 3 mM CaCl<sub>2</sub>.
- 5. Disperse cells by passing through a sterile stainless steel or nylon mesh. Remaining tissue fragments may be disaggregated by addition to fresh collagenase solution and further incubation at 37°C.
- 6. Wash dispersed cells several times by centrifugation in HBSS w/o collagenase.
- 7. Resuspend cell pellet, after the final wash step, in culture medium. Determine viable cell density using a Automated Cell Counter (alternate automated or manual methods may be used).
- 8. Seed cells into culture vessels containing appropriate media.

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### Organ Perfusion

- 1. Add collagenase to prewarmed (37°C) HBSS with calcium and magnesium. Addition of 3 mM  $CaCl_2$  increases the efficiency of dissociation.
- 2. Perfuse organ at preoptimized rate for the particular organ.
- 3. Dispersed cells and tissue fragments are separated from larger pieces by passing the perfusate through a sterile stainless steel or nylon mesh. Remaining tissue fragments may be disaggregated by addition to fresh collagenase solution and further incubation at 37°C.
- 4. The steps are the same as for tissue isolation 6-8.

MCE has not independently confirmed the accuracy of these methods. They are for reference only.

### **REFERENCES**

[1]. Hamzeh Alipour, et al. Therapeutic applications of collagenase (metalloproteases): A review. Asian Pac J Trop Biomed. 2016, 6, 11.

Caution: Product has not been fully validated for medical applications. For research use only.

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