MCE USA

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Protein A/G Magnetic Beads

1 Contents

| Cat. No. | Product Name | Package |
|---------------|----------------------------|----------|
| HY-K0202-1 mL | Protein A/G Magnetic Beads | 1 mL |
| HY-K0202-5 mL | Protein A/G Magnetic Beads | 1 mL × 5 |

2 General Information

Protein A/G Magnetic Beads provide a fast and convenient method for magnetic isolation of proteins using affinity binding. MCE Protein A/G Magnetic Beads are typically used for isolating antibodies from serum, cell culture supernatant or ascites and for Immunoprecipitation and Co-Immunoprecipitation of antiqens from cell or tissue extracts.

The MCE Protein A/G Magnetic Beads contain a recombinant Protein A/G that combines the IgG binding domains of both Protein A and Protein G. Protein A/G contains four Fc-binding domains from Protein A and two from Protein G, making it a more general and convenient tool for investigating and purifying immunoglobulins.

3 Characteristics

| Composition | Recombinant Protein A/G monolayer covalently coupled to a blocked magnetic bead surface | |
|--------------------|---|--|
| Magnetization | Superparamagnetic | |
| Mean Diameter | 200 nm | |
| Bead Concentration | 10 mg/mL | |
| Binding Capacity | 0.7 mg/mL | |
| | | |

4 General Protocol

Recommended Buffer

| Binding/Wash Buffer | PBST : 1× PBS + 0.5% Tween-20, pH 7.4 |
|---------------------|---------------------------------------|
| Elution Buffer | 0.15 M Glycine, pH 2.5-3.1 |

- 1. Preparation of Magnetic Beads
- 1.1 Resuspend the Magnetic Beads in the vial (tilt and rotate for 2 minutes or gently pipette for 10 times).
- 1.2 Transfer 25-50 μ L of Protein A/G Magnetic Beads into a 1.5 mL tube (Transfer amount may be adjusted as required).

- 1.3 Add 400 μ L of binding/wash buffer to the beads and gently pipette to mix. Place the tube into a magnetic stand to collect the beads against the side of the tube (Hereinafter referred to as magnetic separation). Remove and discard the supernatant. Repeat this step for 2 times.
- 2. Binding of Antibody
- 2.1 Dilute antibody (Ab) to the final concentration of 5-50 μ g/mL with binding/wash buffer. The optimal amount of Ab may be adjusted as required.
- 2.2 Add 400 μ L of diluted Ab to the Protein A/G Magnetic Beads. Rotate tube for 30 minutes at room temperature or 2 hours at 4°C.
- 2.3 Perform magnetic separation. Transfer the supernatant into a new tube for further analysis, if desired. The supernatant is the non-binding fraction.
 2.4 Add 400 μ L of binding/wash buffer to the beads and gently pipette to mix. Place the tube into a magnetic stand to collect the beads against the side of the tube. Remove and discard the supernatant. Repeat this step for 4 times.

Note: During the binding process, it won't affect the result if magnetic beads occasionally aggregated together.

- 3. Immunoprecipitation of Target Antigen
- 3.1 Remove the tubes from the magnetic separator and add your sample containing the antigen (Ag) (typically 5-50 μg in 400 μL lysis buffer) and gently pipette to resuspend the Protein A/G Magnetic Beads-Ab complex.
- 3.2 Incubate with rotation for 30 minutes at room temperature or 2 hours at 4°C to allow Ag to bind to the Protein A/G Magnetic Beads-Ab complex.

 Note: Depending on the affinity of antibody, it may be necessary to increase the incubation time for optimal binding.
- 3.3 Perform magnetic separation. Remove and discard the supernatant.
- 3.4 Wash the Magbeads-Ab-Ag complex 5 times using 400 μ L binding/wash buffer for each wash. Perform magnetic separation between each wash, remove supernatant and resuspend by gentle pipetting.
- 3.5 Resuspend the Protein A/G Magnetic Beads-Ab-Ag complex in 400 μ L binding/wash buffer and transfer the bead suspension into a clean tube. This is recommended to avoid co-elution of the proteins bound to the tube wall.

4. Elution

This is a non-denaturation elution method.

4.1 Perform magnetic separation and remove the supernatant. Add 400 μL of binding/wash buffer into the tube and rotate for 5 minutes. Perform magnetic separation for 1 minute and remove the supernatant. Then add

- 25-50 μL elution buffer into the tube with magnetic beads-Ab-Ag complex, rotate for 5 minutes.
- 4.2 Perform magnetic separation, collect the supernatant.
- 4.3 The final solution can be used as samples for denaturing SDS-PAGE. Or the elution can be adjusted to neutral pH with neutralization buffer immediately and used for further analysis.

5 Storage

Stored at 4°C, and is stable for up to 2 years.

6 Precautions

- 1. The pH of Protein A/G Magnetic Beads is 6-8.
- 2. Do not centrifuge, dry or freeze the magnetic beads.
- 3. This product is for R&D use only, not for drug, house hold, or other uses. Please consult the Material Safety Data Sheet for information regarding hazards and safe handling practices.

7 Troubleshooting

| Problem | Possible Cause | Solution |
|---|---|---|
| Low binding efficiency of antibody and magnetic beads | The binding efficiency of magnetic beads and antibody depends on the origin and subtype of the antibody | Check the affinity of antibody and the protein A/G matrix in the appendix |
| | The antibody subtype and protein A/G matrix shows low affinity | Elongate the incubation time of antibody and magnetic beads |
| | | Increase the pH vaule of binding buffer (8-9) and reduce the ion strength (25-100 mM NaCl) |
| Magnetic beads aggregated | Magnetic beads were frozen or centrifuged | Handle the beads as directed in the instructions |
| | Buffer was incompatible with magnetic beads | |
| Multiple nonspecific bands | Nonspecific proteins bound to the magnetic beads | Add 50-350 mM of NaCl to the binding/washing and elution buffers |
| Protein not eluted | Elution conditions were too mild | Increase incubation time with elution buffer or use more stringent elution buffer |
| Low amount of protein recovered | The protein degraded | Add protease inhibitors (e.g., HY-K0010 or HY-K0011) |
| | Not enough magnetic beads were used | Increase the amount of magnetic beads used for capture |
| | Sample had an insufficient amount of target protein | Increase amount of antigen sample |

Appendix: Binding Affinity of Protein A/G for Different Antibodies and Subtypes

| Species | Antibody Subtype | Protein A/G |
|------------|----------------------|--------------------|
| Human | Total IgG | ++++ |
| | IgG1, IgG2 | ++++ |
| | IgG3 | ++++ |
| | IgG4 | ++++ |
| | IgM | + |
| | IgD | - |
| | IgA | + |
| | IgA1, IgA2 | + |
| | IgE | +++ |
| | Fab | + |
| | ScFv | + |
| Mouse | Total IgG | ++++ |
| | IgM | - |
| | IgG1 | +++ |
| | IgG2a | +++ |
| | IgG2b | +++ |
| | IgG3 | +++ |
| Rat | Total IgG | +++ |
| | IgG1 | +++ |
| | IgG2a | ++++ |
| | IgG2b | + |
| | IgG2c | ++++ |
| Cow | Total IgG | ++++ |
| | IgG1 | ++++ |
| | IgG2 | ++++ |
| Goat | Total IgG | ++++ |
| | IgG1 | ++++ |
| | IgG2 | ++++ |
| Sheep | Total IgG | ++++ |
| | IgG1 | ++++ |
| | IgG2 | ++++ |
| Horse | Total IgG | ++++ |
| | IgG(ab), IgG(c) | + |
| | IgG(T) | ++++ |
| Rabbit | Total IgG | ++++ |
| Guinea Pig | Total IgG | ++++ |
| Hamster | Total IgG | +++ |
| Pig | Total IgG | ++++ |
| Donkey | Total IgG | ++++ |
| Cat | Total IgG | ++++ |
| Dog | Total IgG | ++++ |
| Monkey | Total IgG | ++++ |
| Chicken | Total IgG | - |
| Notes: | + weak binding | +++ medium binding |
| | +++++ strong binding | - no biding |

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