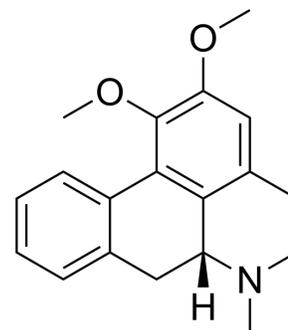


Nuciferine

Cat. No.:	HY-N0049		
CAS No.:	475-83-2		
Molecular Formula:	C ₁₉ H ₂₁ NO ₂		
Molecular Weight:	295.38		
Target:	5-HT Receptor; Dopamine Receptor; Parasite		
Pathway:	GPCR/G Protein; Neuronal Signaling; Anti-infection		
Storage:	Powder	-20°C	3 years
		4°C	2 years
	In solvent	-80°C	2 years
		-20°C	1 year



SOLVENT & SOLUBILITY

In Vitro	DMSO : 5 mg/mL (16.93 mM; ultrasonic and warming and heat to 60°C)				
		Solvent Concentration	Mass 1 mg	5 mg	10 mg
	Preparing Stock Solutions	1 mM	3.3855 mL	16.9273 mL	33.8547 mL
		5 mM	0.6771 mL	3.3855 mL	6.7709 mL
10 mM		0.3385 mL	1.6927 mL	3.3855 mL	
Please refer to the solubility information to select the appropriate solvent.					
In Vivo	1. Add each solvent one by one: 10% DMSO >> 40% PEG300 >> 5% Tween-80 >> 45% saline Solubility: ≥ 1.11 mg/mL (3.76 mM); Clear solution 2. Add each solvent one by one: 10% DMSO >> 90% corn oil Solubility: ≥ 1.11 mg/mL (3.76 mM); Clear solution				

BIOLOGICAL ACTIVITY

Description	Nuciferine is an antagonist at 5-HT _{2A} (IC ₅₀ =478 nM), 5-HT _{2C} (IC ₅₀ =131 nM), and 5-HT _{2B} (IC ₅₀ =1 μM), an inverse agonist at 5-HT ₇ (IC ₅₀ =150 nM), a partial agonist at D ₂ (EC ₅₀ =64 nM), D ₅ (EC ₅₀ =2.6 μM) and 5-HT ₆ (EC ₅₀ =700 nM), an agonist at 5-HT _{1A} (EC ₅₀ =3.2 μM) and D ₄ (EC ₅₀ =2 μM) receptor.			
IC₅₀ & Target	5-HT _{2C} Receptor 131 nM (IC ₅₀)	Schistosome	5-HT ₇ Receptor 150 nM (IC ₅₀)	5-HT _{2A} Receptor 478 nM (IC ₅₀)
	5-HT _{2B} Receptor 1 μM (IC ₅₀)	5-HT ₆ Receptor 700 nM (EC ₅₀)	5-HT _{1A} Receptor 3.2 μM (EC ₅₀)	D ₂ Receptor 64 nM (EC ₅₀)

	D ₄ Receptor 2 μM (EC50)	D ₅ Receptor 2.6 μM (EC50)
In Vitro	<p>Nuciferine is a partial agonist at DD₂ receptor with an activity (E_{max}=67% of dopamine) similar to aripiprazole (E_{max}=50% of dopamine). In line with its partial agonist activity, Nuciferine inhibited dopamine-induced activation of G_i with a potency similar to clozapine (Nuciferine K_B=62 nM; Clozapine K_B=20 nM) as determined via Schild regression analysis^[1]. The natural product Nuciferine acts as an effective inhibitor of adult worm motility. Nuciferine is effective at inhibiting both basal and 5-HT evoked motility of adult schistosomes. Nuciferine inhibits Sm.5HTR_L and schistosomule with 0.24±0.04 and 0.62±0.22 μM, respectively^[2].</p> <p>MCE has not independently confirmed the accuracy of these methods. They are for reference only.</p>	
In Vivo	<p>In rodent models relevant to antipsychotic drug action, Nuciferine blocks head-twitch responses and discriminative stimulus effects of a 5-HT_{2A} agonist, substituted for clozapine discriminative stimulus, enhanced amphetamine induced locomotor activity, inhibited phencyclidine (PCP)-induced locomotor activity, and rescued PCP-induced disruption of prepulse inhibition without induction of catalepsy. In the presence of 1 or 3 mg/kg Nuciferine, cumulative PCP doses produce similar substitution to PCP alone. In the clozapine-trained animals, a dose-dependent substitution for 1.25 mg/kg clozapine is seen at 10 mg/kg Nuciferine (80.63% drug lever responding), with an ED₅₀ value of 5.42 mg/kg (95% CI 3.09-9.48 mg/kg) while the lower doses tested (0.1 mg/kg-3 mg/kg) fails to produce substitution for clozapine's discriminative cue. In addition to a high percentage of responding on the clozapine-appropriate lever, 10 mg/kg Nuciferine also produces significant rate suppression as compared to vehicle control points (p<0.001)^[1].</p> <p>MCE has not independently confirmed the accuracy of these methods. They are for reference only.</p>	

PROTOCOL

Kinase Assay ^[1]	<p>For affinity determination, Nuciferine is subjected to primary radioligand binding assays tested at a single 10 μM concentration to displace 50% of the radioligand at a given receptor target. If a more than 50% of the radioligand is displaced, Nuciferine is selected for a secondary binding assay tested at 11 concentrations in triplicate in competition with the radioligand to generate an IC₅₀ and K_i. Binding assays are performed in 96-well plates with 125 μL per well in appropriate binding buffer using radioligand at or near the K_d. Plates are incubated at room temperature in the dark for 90 min. Reactions are stopped by vacuum filtrations onto 0.3% polyethyleneimine soaked 96-well filter mats using a 96-well Filtermate harvester, followed by at least three washes of cold wash buffer. Scintillation cocktail is melted onto dried filters and radioactivity is counted using a Wallac Trilux Microbeta^[1].</p> <p>MCE has not independently confirmed the accuracy of these methods. They are for reference only.</p>
Cell Assay ^[1]	<p>Cells are plated into 48-well plates one day before uptake is performed. Cells are washed with 0.5 mL uptake buffer (4 mM Tris, 6.25 mM HEPES, 120 mM NaCl, 5 mM KCl, 1.2 mM CaCl₂, 1.2 mM MgSO₄, 5.6 mM D-glucose, 1.7 mM ascorbic acid, and 1 μM pargyline, pH 7.4). Cells are incubated with 225 μL uptake buffer with or without the indicated concentration of Nuciferine for 15 minutes. After incubation, 25 μL uptake buffer containing ³H-DA and DA is added for a final concentration of 20 nM ³H-DA and 1 μM DA. Cells are incubated at 37°C for 20 minutes or for the time indicated. Nonspecific uptake is determined in the presence of 10 μM nomifensine. Uptake is terminated by aspirating uptake buffer and washing each well twice with 0.5 mL ice-cold uptake buffer. Cells are lysed in 0.1 N NaOH and transferred to vials containing 3 mL scintillation cocktail. Radioactivity is quantitated using a Beckman LS6500 counter. Data are analyzed in Graph Pad Prism 5.0^[1].</p> <p>MCE has not independently confirmed the accuracy of these methods. They are for reference only.</p>
Animal Administration ^[1]	<p>Mice^[1]</p> <p>Adult male NIH Swiss mice weighing approximately 25 g are used. Mice are injected with either Nuciferine (1, 3, or 10 mg/kg, i.p.) or vehicle, n=4 mice/condition. Fifteen minutes later, mice are injected with 1 mg/kg DOI (i.p.) and immediately placed in an observation chamber (new cage without bedding). Head-twitches (operationally defined as a rapid rotational jerk of the head that can be distinguished from species-appropriate grooming or scratching behaviors) are counted for 20 minutes in 5 minute bins. For the time-course study, mice are pretreated with 3.0 mg/kg Nuciferine (i.p.) at 60, 45, 30, 15, or 0 minutes (co-injection) prior to the 1.0 mg/kg DOI (i.p.) injection, and head-twitches are counted as described above. In one experiment, mice (n=4 per condition) are pretreated with an injection (s.c.) of 3.0 mg/kg Nuciferine or vehicle 15 minutes</p>

prior to 1.0 mg/kg DOI injection (i.p.) and head-twitches are counted as described above. All experiments are performed by 3 observers, with 2 observers blinded to the experimental conditions which are evenly distributed. Power analyses are performed with the resulting data. The two highest doses of Nuciferine tested (10 and 3 mg/kg), had 0.96 and 0.88 power to detect significance ($\alpha=0.05$). As these experiments are performed blinded and in distinct mice, further replication is not performed.

MCE has not independently confirmed the accuracy of these methods. They are for reference only.

CUSTOMER VALIDATION

- Int Immunopharmacol. 2023 Apr 29;119:110204.
- J Funct Foods. August 2022, 105182.
- Nutr Metab. 2021 Feb 18;18(1):20.
- Mol Pharmacol. 2023 Nov;104(5):230-238.

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REFERENCES

- [1]. Farrell MS, et al. In Vitro and In Vivo Characterization of the Alkaloid Nuciferine. PLoS One. 2016 Mar 10;11(3):e0150602.
- [2]. Chan JD, et al. Pharmacological profiling an abundantly expressed schistosome serotonergic GPCR identifies nuciferine as a potent antagonist. Int J Parasitol Drugs Drug Resist. 2016 Dec;6(3):364-370.

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