

Human Kidney Tubular Organoid Kit

1 Contents

Cat.No.	Component	HY-K6116-100 mL	HY-K6116-500 mL
HY-K6116-A	Human Kidney Tubular Organoid Basal Medium A	100 mL	500 mL
HY-K6116-B	Human Kidney Tubular Organoid Supplement B (50x)	1 mL × 2	10 mL
HY-K6116-C	Human Kidney Tubular Organoid Supplement C (250x)	0.4 mL	1 mL × 2

2 Introduction

MCE Human Kidney Tubular Organoid Kit contains Kidney Tubular Organoid Basal Medium A, Kidney Tubular Organoid Supplement B (50x), Kidney Tubular Organoid Supplement C (250x). This product can be used to efficiently construct human kidney tubular cancer organoid.

The structure and function of the kidney tubular organoid (tubuloids) are similar to that of real kidney tubules. Kidney tubular organoid can be used for many applications such as organ development, disease modeling, drug screening, nephrotoxic testing, and regenerative therapy.

3 Operation Instructions

1. Preparation of complete culture medium for kidney tubular

Prepare complete medium for kidney tubular organoid according to the following components, mix thoroughly and set aside on ice.

Reagents	10 mL	20 mL	50 mL	Final concentration
Human Kidney Tubular Organoid Basal Medium A	9.76 mL	19.52 mL	48.6 mL	1x
Human Kidney Tubular Organoid Supplement B (50x)	200 µL	400 µL	1 mL	1x
Human Kidney Tubular Organoid Supplement C (250x)	40 µL	80 µL	200 µL	1x

2. Extraction of cells from primary tissues

- Soak freshly extracted primary tissue using pre-cooled primary tissue storage solution and store them temporarily in a 4°C refrigerator.
- Rinse with Human Kidney Tubular Organoid Basal Medium A or PBS to remove non-epithelial tissue components such as fat or muscle under guaranteed aseptic condition.
- Use sterile scissors to divide the rinsed primary tissue into the smallest possible pieces (approximately 2 mm in diameter) in a cell culture dish, ensuring they are small enough to pass through the tip of a 10 mL pipette. Transfer them to a 15 mL conical tube containing 10 mL of cold Human Kidney Tubular Organoid Basal Medium A with 1% FBS.
- Prepare Human Kidney Tubular Organoid Basal Medium A with 10% FBS (10% FBS medium). Coat the inner surface of pipette tips with 10% FBS medium before use to avoid the adherence of the samples on the pipette wall.

- e. Wash the samples by pipetting with a 10 mL pipette at least ten times. Stand the tube and let the samples settle by gravity.
- f. Aspirate the supernatant with a 10 mL pipette. Add an appropriate amount of pre-warmed Tissue Digestion Solution, preferably not exceeding two-thirds of the volume of the conical tube. Incubate the conical tube on a horizontal shaker at 37°C for no more than 30 min.
- g. Check digestion under the microscope if the duct structure appears during digestion, and once it appears, stop digestion by adding FBS to a final concentration of 2% and pipetting gently up and down.
- h. Stand the tube and let the samples settle by gravity for 1-2 min. Collect the supernatant into a new tube.
- i. Add 10 mL of Human Kidney Tubular Organoid Basal Medium A. Stand the tube and let the samples settle by gravity for 1-2 min, and collect the supernatant into a new tube. Collect the cell precipitate after centrifugation at 300 g for 3 min at 4°C using a cryogenic centrifuge.
- j. Resuspend the collected cell precipitate by adding 10 mL of Human Kidney Tubular Organoid Basal Medium A, and then collect the cell precipitate by centrifugation at 300 g for 3 min in a low-temperature centrifuge at 4°C. Repeat this step twice.

3. Construction of organoid

- a. Resuspend the collected primary cells in MCE Basement Membrane Matrix on ice. Approximately 50-200 ducts should be plated in 50 µL Basement Membrane Matrix. It is recommended that 100% MCE Basement Membrane Matrix be used to resuspend tumor cells. If dilution is required, please ensure that the ratio of MCE Basement Membrane Matrix volume to the volume of organoid medium used for dilution is greater than 2:1.
- b. Quickly inject MCE Basement Membrane Matrix with the cell suspension into the bottom of the 24-well cell culture plate, avoiding air bubbles as much as possible, injecting 25-35 µL of suspension per well. The cell culture plate is then incubated in a 37°C, 5% CO₂ incubator for 15-30 min until gelling.
- c. Inject 500 µL of kidney tubular complete culture medium at the edge of each well slowly to avoid disrupting the existing gel structure after gelling. Then place the cell culture plate back into the incubator at 37°C, 5% CO₂.
- d. Replace 500 µL of pre-warmed kidney tubular complete culture medium volume at 37°C every 3 days. Kidney tubular organoid can be observed in 5-8 days.

4. Organoid passages

- a. It is recommended to aspirate the upper medium and add 500 µL of Kidney Tubular Organoid Basal Medium A. Use pipette tip to blow to peel the contents of the cell culture wells out of the plate and transfer them to a 1.5 mL EP tube.
- b. Blow gently until the kidney tubular organoid is separated from MCE Basement Membrane Matrix using a pipette tip. Then collect the precipitate by centrifugation at 250-300 g for 3 min at room temperature.
- c. Add 1 mL of Human Kidney Tubular Organoid Basal Medium A and resuspend and gently blow well until the organoids are dispersed into fragments. If the organoid is difficult to be blown into pieces, use an appropriate amount of organoid digestion solution in 37°C incubator to digest the organoid until it is dispersed into cell clusters containing 10-50 cells. The digestion time should be limited to 3 min or less. The digestion is then terminated by adding 1 mL Human Kidney Tubular Organoid Basal Medium A.
- d. Centrifuge at 200-250 g for 3 min at room temperature. After centrifugation, the supernatant was discarded and washed 1-2 times with Kidney Tubular Organoid Basal Medium A or PBS and then prepared for use.
- e. Resuspend the collected primary cells in MCE Basement Membrane Matrix on ice. Approximately 40-200 ducts should be plated in 50 µL Basement Membrane Matrix. It is recommended that 100% MCE Basement Membrane Matrix be used to resuspend tumor cells. If dilution is required, please ensure that the ratio of MCE Basement Membrane Matrix volume to the volume of organoid medium used for dilution is greater than 2:1.
- f. Quickly inject MCE Basement Membrane Matrix with the cell suspension into the bottom of the 24-well cell culture plate, avoiding air bubbles as much as possible, injecting 25-35 µL of suspension per well. The cell culture plate is then incubated in a 37°C, 5% CO₂ incubator for 15-30 min until gelling.
- g. Inject 500 µL of kidney tubular complete culture medium at the edge of each well slowly to avoid disrupting the existing gel structure after gelling. Then place the cell culture plate back into the incubator at 37°C, 5% CO₂.
- h. Replace 500 µL of pre-warmed kidney tubular complete culture medium volume at 37°C every 3-4 days.

4 Storage

Individual Components	Human Kidney Tubular Organoid Basal Medium A	4°C, 1 year.
	Human Kidney Tubular Organoid Supplement B (50x)	-20°C, 1 year. Avoid repeated freeze/thaw cycles.
	Human Kidney Tubular Organoid Supplement C (250x)	-20°C, 1 year. Avoid repeated freeze/thaw cycles.
Complete Culture Medium	Human Kidney Tubular Organoid Complete Culture Medium	4°C, 2 weeks or -20°C, 3 months. Avoid repeated freeze/thaw cycles.

Note: It is recommended that individual components be formulated for use immediately after thawing. It is better to prepare complete culture medium fresh before the experiment, otherwise please make aliquots for freezing.

5 Precautions

1. Primary tissue cells need to be kept sterile when extracted from primary tissue to avoid contamination from subsequent experiments.
2. Observe the fragmentation status of the organoid during passaged digestion, and terminate the digestion when small cell clusters (10-50 cells) appear to avoid prolonging the subsequent growth viability of the organoid.
3. Operations involving MCE Basement Membrane Matrix need to be kept at low temperature throughout. MCE Basement Membrane Matrix should be injected rapidly into the cell culture wells after resuspension with the cells, while avoiding air bubbles.
4. This product is for R&D use only, not for drug, household, or other uses.
5. For your safety and health, please wear a lab coat and disposable gloves to operate.